

Organoid media formulation #6

The following components are required for media preparation:

| Item | Vendor | Catalog # | Size | Website | |
|---|---|-------------------|----------|------------------|--|
| Advanced DMEM:F12 | Thermo Fisher | 12634028 | 500 mL | thermofisher.com | |
| HEPES | Thermo Fisher | 15630080 | 100 mL | thermofisher.com | |
| B-27 Supplement | Thermo Fisher | 17504-044 | 10 mL | thermofisher.com | |
| L-Glutamine | ATCC | 30-2214™ | 100 mL | atcc.org | |
| Dimethyl sulfoxide (DMSO) | ATCC | 4-X TM | 25 mL | atcc.org | |
| EGF | Bio-techne | 236-EG | 200 µg | bio-techne.com | |
| Gastrin | Bio-techne | 3006 | 1 mg | bio-techne.com | |
| FGF-10 | Bio-techne | 345-FG | 2x 25 μg | bio-techne.com | |
| Nicotinamide | LKT Labs | N3310 | 50 g | lktlabs.com | |
| N-acetyl cysteine | LKT Labs | A0918 | 10 g | lktlabs.com | |
| TGF-beta 1 | Bio-techne | 240-B-010 | 10 µg | bio-techne.com | |
| HA-R-Spondin1-Fc 293T (RSPO1) Conditioned Media | For each 500 mL of complete organoid media, 50 mL of RSPO1 conditioned media is required. Refer to vendors instructions to prepare conditioned medium from Trevigen Cultrex® HA-R-Spondin1-Fc 293T Cells (Trevigen Cat # 3710-001-01). The protocol for cell culture and conditioned medium generation is available at: https://trevigen.com/docs/protocol/protocol 3710-001-01.pdf | | | | |
| CRL-2647 L Wnt-3A Conditioned Media | For each 500 mL of complete organoid media, 250 mL of WNT3A conditioned media is required. Refer to the product sheet for instructions to prepare conditioned medium from ATCC CRL-2647 L Wnt-3A cells. The protocol for cell culture and conditioned medium generation is available at: https://www.atcc.org/~/ps/CRL-2647.ashx | | | | |

Refer to the manufacturer of individual components for important safety and handling considerations.

Media preparation procedure

- 1. Thaw B-27 and L-Glutamine on ice or in a refrigerator at 2-8°C. Aliquot into working volumes and freeze. Do not re-freeze/thaw multiple times.
- 2. Briefly centrifuge the vials containing the EGF, Gastrin and FGF-10 (2 vials) to ensure the material is at the bottom of the vial.
- 3. Aseptically reconstitute the following components: EGF, FGF-10, and Gastrin according to the manufacturer's instructions in the recommended buffer listed in the table below. We recommend incubating in buffer for 15 minutes at room temperature.

| Item | Size | Buffer | Volume of Buffer | Final Concentration |
|------------------|-----------|---------------------------|---------------------|---------------------|
| EGF | 200 µg | Advanced DMEM:F12 | 2.0 mL | 100 μg/mL |
| Gastrin | 1 mg | Advanced DMEM:F12 | 4.7 mL | 100 μM |
| FGF-10 (2 vials) | 2 x 25 µg | Advanced DMEM:F12 | 2x 0.25 mL | 100 μg/mL |
| TGF-B1 | 10 µg | 4 mM sterile filtered HCI | 1.0 mL | 10 μg/mL |



4. Aseptically weigh and prepare working solutions of Nicotinamide and N-Acetyl Cysteine in the recommended buffer. If N-Acetyl Cysteine is difficult to dissolve, periodic vortexing and incubation in a 37.0°C water bath can help the material enter solution.

| Item | Weight | Buffer | Volume of Buffer | Final Concentration |
|----------------------|--------|-------------------|------------------|---------------------|
| Nicotinamide | 5 g | Advanced DMEM:F12 | 41.0 mL | 1 M |
| N-Acetyl Cysteine | 2.5 g | Advanced DMEM:F12 | 61.0 mL | 250 mM |

5. Prepare the complete growth medium formulation (makes 500 mL):

| Item | Volume | Final Concentration |
|-------------------------|-------------|---------------------|
| Advanced DMEM:F12 | 171.5 mL | N/A |
| HEPES | 5.0 mL | 10 mM |
| L-Glutamine | 5.0 mL | 2 nM |
| B-27 | 10.0 mL | 1X |
| EGF | 0.25 mL | 50 ng/mL |
| FGF-10 (2 vials) | 2 x 0.25 mL | 100 ng/mL |
| Gastrin | 50.0 μL | 10 nM |
| Nicotinamide | 5.0 mL | 10 mM |
| N-Acetyl Cysteine | 2.5 mL | 1.25 mM |
| RSPO1 conditioned media | 50 mL | 10% |
| WNT3A conditioned media | 250 mL | 50% |

- 6. <u>For the first 2-3 weeks of culture only</u>, supplement the complete media with the prepared TGF-B1 to a final concentration of 5 ng/mL by adding 1 μL per 2 mL complete growth medium. After 2-3 weeks TGF-B1 is no longer included in the complete growth medium.
- 7. Once prepared, store complete medium at 2-8°C in the dark. Do not freeze and avoid extended exposure to light. Label with date of preparation and discard after 30 days.
- 8. When using the medium during culture, only warm the volume required.
- 9. Refer to the manufacturer's documentation for appropriate storage conditions and stability of individual components once reconstituted.

Notes

- Purity and activity levels of the various components can change from lot-lot. Refer to the manufacturer's Certificates of Analysis to ensure equivalent quality when using a new lot of material.
- We do not recommend deviating from the formulation or substituting components from different vendors.



• We recommend that solutions are prepared on the same day they are used. If the solutions must be stored, aliquot and freeze at -80°C or below and use within 30 days. Once reconstituted the components will lose activity over time and this can negatively affect performance of the medium.

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