



Rhodobacter capsulatus (Molisch) Imhoff et al.

17018™

Description

Strain designation: ATH 2.3.5

Deposited As: *Rhodopseudomonas capsulatus* (Molisch) van Niel

Type strain: No

Storage Conditions

Product format: Freeze-dried

Intended Use

This product is intended for laboratory research use only. It is not intended for any animal or human therapeutic use, any human or animal consumption, or any diagnostic use.

BSL 1

ATCC determines the biosafety level of a material based on our risk assessment as guided by the current edition of *Biosafety in Microbiological and Biomedical Laboratories (BMBL)*, U.S. Department of Health and Human Services. It is your responsibility to understand the hazards associated with the material per your organization's policies and procedures as well as any other applicable regulations as enforced by your local or national agencies.

ATCC highly recommends that appropriate personal protective equipment is always

used when handling vials. For cultures that require storage in liquid nitrogen, it is important to note that some vials may leak when submersed in liquid nitrogen and will slowly fill with liquid nitrogen. Upon thawing, the conversion of the liquid nitrogen back to its gas phase may result in the vial exploding or blowing off its cap with dangerous force creating flying debris. Unless necessary, ATCC recommends that these cultures be stored in the vapor phase of liquid nitrogen rather than submersed in liquid nitrogen.

Certificate of Analysis

For batch-specific test results, refer to the applicable certificate of analysis that can be found at www.atcc.org.

Growth Conditions

Medium:

ATCC Medium 112: Van Niel's yeast agar

Temperature: 30°C

Atmosphere: Anaerobic

Handling Procedures

- 1. Put 6 to 8 ml of medium #112 into a 13 x 100 mm screw cap test tube (small). Add cysteine (3.0% stock concentration, 2 ml/100 ml medium) and then fill the test tube to capacity with additional #112 broth. Seal the test tube with a screw cap.**
- 2. Let the tube sit at room temperature for 30 minutes before inoculating it with the rehydrated culture.**
- 3. Aseptically take 0.5 ml of the pre-reduced medium and rehydrate the entire**

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pellet.

4. Transfer the rehydrated pellet back into the screw cap test tube and close tightly. (The test tube should be filled to capacity).

5. Incubate the culture at 30°C under a tungsten lamp.

6. Once growth has been established (three to six days), the culture should be transferred to fresh broth. If a large inoculum is used (1020%), the medium does not need to be pre-reduced.

7. When examined microscopically, the cells are long, thin rods that occur singly, in pairs, and in clumps. The cells are motile.

Notes

This culture is tolerant to oxygen; therefore, strictly anoxic conditions are not required when rehydrating the freeze-dried pellet or transferring the organism.

This culture is able to grow aerobically on nutrient agar in the dark. Colonies are circular, smooth, entire, and violet.

Material Citation

If use of this material results in a scientific publication, please cite the material in the following manner: *Rhodobacter capsulatus* (Molisch) Imhoff et al. (ATCC 17018)

References

References and other information relating to this material are available at www.atcc.org.

Warranty

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