



Homalogastra setosa Kahl

50260™

Description

Strain designation: RV

Deposited As: *Homalogastra setosa* Kahl

Type strain: No

Storage Conditions

Product format: Dried

Intended Use

This product is intended for laboratory research use only. It is not intended for any animal or human therapeutic use, any human or animal consumption, or any diagnostic use.

BSL 1

ATCC determines the biosafety level of a material based on our risk assessment as guided by the current edition of *Biosafety in Microbiological and Biomedical Laboratories (BMBL)*, U.S. Department of Health and Human Services. It is your responsibility to understand the hazards associated with the material per your organization's policies and procedures as well as any other applicable regulations as enforced by your local or national agencies.

ATCC highly recommends that appropriate personal protective equipment is always

used when handling vials. For cultures that require storage in liquid nitrogen, it is important to note that some vials may leak when submersed in liquid nitrogen and will slowly fill with liquid nitrogen. Upon thawing, the conversion of the liquid nitrogen back to its gas phase may result in the vial exploding or blowing off its cap with dangerous force creating flying debris. Unless necessary, ATCC recommends that these cultures be stored in the vapor phase of liquid nitrogen rather than submersed in liquid nitrogen.

Certificate of Analysis

For batch-specific test results, refer to the applicable certificate of analysis that can be found at www.atcc.org.

Growth Conditions

Medium:

ATCC Medium 802: Sonneborn's Paramecium medium

Instructions for complete medium:

ATCC Medium 802 inoculated with *Klebsiella pneumoniae* subsp. *pneumoniae* (ATCC® 700831™).

Temperature: 25°C

Culture system: Xenic

Incubation: grown with *Enterobacter aerogenes* ATCC 13048 and mixed bacteria

Handling Procedures

Establishing Cultures from Dried State

This strain comes dried on shredded filter paper. Dried samples can remain at room

temperature for up to one week. If the culture will not be rehydrated within that period, store at 5°C until processed.

1. To rehydrate an ampule, aseptically add 0.5 ml of sterile ATCC medium 802 or sterile distilled water to the inner shell vial. Aseptically remove the filter paper pellet with a pair of forceps and transfer it to a T-25 flask containing 10 ml of ATCC medium 802 bacterized with *Klebsiella pneumoniae* subsp. *pneumoniae* (ATCC® 700831).
2. Using a Pasteur pipette, aseptically transfer the remainder of the liquid from the vial to the T-25 flask. Agitate the flask to break apart the filter paper pellet, and incubate the culture at 25°C with the cap screwed on tightly.
3. On the following day, vigorously agitate the culture and aseptically transfer a 0.5-ml aliquot to 10 ml of bacterized ATCC medium 802. The subculture will have almost completely encysted within 3-4 days.

Culture maintenance: Subculture every two to three weeks to a fresh T-25 flask of bacterized medium in the following manner:

1. Vigorously agitate the flask and aseptically transfer 0.5 ml from a growing culture to a T-25 tissue culture flask containing 10.0 ml of ATCC medium 802 bacterized with *Klebsiella pneumoniae* subsp. *pneumoniae* (ATCC® 700831)
2. Incubate flask at 25°C with the cap screwed on tightly.

Reagents for cryopreservation:

Cryoprotective Solution

DMSO, 1.5 ml

Fresh growth medium w/o bacteria, 8.5 ml

Cryopreservation:

1. Mix the components in the order listed. When the medium is added to the DMSO the solution will warm up due to chemical heat.
2. Harvest cells from a culture that is at or near peak density by filtration and centrifugation at 800 x g for 5 min.
3. Adjust the concentration of cells at least 2×10^6 /ml in fresh medium.
4. Mix the cell preparation and the cryoprotective solution in equal portions.
5. Dispense in 0.5 ml aliquots into 1.0 - 2.0 ml sterile plastic screw-capped cryules (special plastic vials for cryopreservation).
6. Place vials in a controlled rate freezing unit. From room temperature cool at -1°C/min to -40°C. If freezing unit can compensate for the heat of fusion,

maintain rate at -1 C/min through heat of fusion. At -40°C plunge ampules into liquid nitrogen. Alternatively, place the vials in a Nalgene 1°C freezing apparatus. Place the apparatus at -80°C for 1.5 to 2 hours and then plunge ampules into liquid nitrogen. (The cooling rate in this apparatus is approximately -1°C/min.)

7. Ampules are stored in either the vapor or liquid phase of a nitrogen refrigerator.
8. To establish a culture from the frozen state place the vial in a 35°C water bath. Immerse the vial to a level just above the surface of the frozen material. Do not agitate the vial. Immediately after thawing, do not leave in water bath, aseptically remove the contents of the ampule and inoculate into a T-25 tissue culture flask containing 10 ml ATCC medium 802 bacterized with *Klebsiella pneumoniae* subsp. *pneumoniae* (ATCC® 700831).
9. Incubate at 25°C with the cap screwed on tightly.
10. Once the culture is established, vigorously agitate the flask and aseptically transfer 0.5 ml to 10.0 ml of bacterized ATCC medium 802.
11. Follow the protocol for maintenance of culture.

Material Citation

If use of this material results in a scientific publication, please cite the material in the following manner: *Homalogastra setosa* Kahl (ATCC 50260)

References

References and other information relating to this material are available at www.atcc.org.

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