



BAA-1389

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Description

Desulfurispirillum indicum strain S5 is a bacterial type strain that was isolated from Buckingham canal in India.

Strain designation: Strain S5

Deposited As: *Selenospirillum indicus*

Type strain: No

Storage Conditions

Product format: Frozen

Storage conditions: -80°C or colder

Intended Use

This product is intended for laboratory research use only. It is not intended for any animal or human therapeutic use, any human or animal consumption, or any diagnostic use.

BSL 1

ATCC determines the biosafety level of a material based on our risk assessment as guided by the current edition of *Biosafety in Microbiological and Biomedical Laboratories (BMBL)*, U.S. Department of Health and Human Services. It is your responsibility to understand the hazards associated with the material per your organization's policies and procedures as well as any other applicable regulations as enforced by your local or national agencies.

ATCC highly recommends that appropriate personal protective equipment is always used when handling vials. For cultures that require storage in liquid nitrogen, it is important to note that some vials may leak when submerged in liquid nitrogen and will slowly fill with liquid nitrogen. Upon thawing, the conversion of the liquid nitrogen back to its gas phase may result in the vial exploding or blowing off its cap with dangerous force creating flying debris. Unless necessary, ATCC recommends that these cultures be stored in the vapor phase of liquid nitrogen rather than submerged in liquid nitrogen.

Certificate of Analysis

For batch-specific test results, refer to the applicable certificate of analysis that can be found at www.atcc.org.

Growth Conditions

Medium:

ATCC Medium 1439: Methanogenium medium

Temperature: 26°C**Atmosphere:** 80% N₂, 20% CO₂; Anaerobic

Handling Procedures

1. Sterilize the top of the Balch tube by spraying it with 70% ethanol and then flaming the top.
2. If needed exchange the gas in the test tube for 80% N₂ - 20% CO₂.
3. Add 0.1 ml of 1 M selenate and 0.1 ml of 2 M pyruvate per each 10 ml of medium. Add 0.1 ml of reducing agent (3% cysteine, stock solution) per each 10 ml of medium. When reducing media, also add the sterile additions before

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- inoculating. Let the medium sit at room temperature for 1 hour.
4. Allow the frozen vial to thaw under anaerobic conditions. Once thawed, take a gassed 1.0 ml syringe tipped with 22-gauge needle and withdraw the entire contents of the thawed vial and immediately transfer it to the prepared Balch tube of #1439 broth. Mix well.
 5. Plate 0.1 ml on a non-selective medium to check for aerobic and anaerobic contamination.
 6. Incubate tubes and one plate under an anaerobic atmosphere at 26°C. Incubate non-selective plate aerobically at 26°C to check for purity. In two weeks time, there should be good growth in broth. No growth should occur on the aerobic plates.

ANAEROBIC CONDITIONS:

1. Balch tubes (available from Bellco Glass, Vineland, NJ) are specially designed for anaerobic work and use an aluminum crimp cap to hold a rubber stopper in place. Needles can easily be inserted through the stopper, and the tubes can be pressurized to 2 atm. Alternatively, serum vials may be used, or screw cap tubes with butyl rubber stoppers, in the latter case the stopper may be removed and the tube placed under a cannula system that dispenses sterile, oxygen free gas for addition of reducing agents or inoculation.
2. To obtain a fully reduced medium, it is necessary that the medium be anoxic and that a reducing agent be added. Common reducing agents are sodium sulfide, cysteine, dithiothreitol, and titanium citrate.

Notes

Growth is indicated by red sedimentation at the bottom of the tube or reddish brown coloration in the media by the metabolism of selenate to selenium. Selenate is the best electron acceptor indicated by growth, but 1 M nitrate (10 mM in culture) is a secondary electron acceptor. Red coloration does not occur without the selenate substrate. The broth tubes should be 60% filled with media. This organism can survive on the bench top for at least 1 year after inoculation. When transferring this culture, use a 10% inoculum.

Cells are vibroid in shape.

Additional information on this culture is available on the ATCC® web site at www.atcc.org.

Material Citation

If use of this material results in a scientific publication, please cite the material in the following manner: BAA-1389 (ATCC BAA-1389)

References

References and other information relating to this material are available at www.atcc.org.

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