



# *Helicobacter pylori* (Marshall et al.) Goodwin et al.

BAA-945™

## Description

**Strain designation:** Baylor Challenge Strain 100 (BCS 100)

**Type strain:** No

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## Storage Conditions

**Product format:** Frozen

**Storage conditions:** -80°C or colder

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## Intended Use

This product is intended for laboratory research use only. It is not intended for any animal or human therapeutic use, any human or animal consumption, or any diagnostic use.

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## BSL 2

ATCC determines the biosafety level of a material based on our risk assessment as guided by the current edition of *Biosafety in Microbiological and Biomedical Laboratories (BMBL)*, U.S. Department of Health and Human Services. It is your responsibility to understand the hazards associated with the material per your organization's policies and procedures as well as any other applicable regulations as enforced by your local or national agencies.

ATCC highly recommends that appropriate personal protective equipment is always used when handling vials. For cultures that require storage in liquid nitrogen, it is important to note that some vials may leak when submersed in liquid nitrogen and will slowly fill with liquid nitrogen. Upon thawing, the conversion of the liquid nitrogen back to its gas phase may result in the vial exploding or blowing off its cap with dangerous force creating flying debris. Unless necessary, ATCC recommends that these cultures be stored in the vapor phase of liquid nitrogen rather than submersed in liquid nitrogen.

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## Certificate of Analysis

For batch-specific test results, refer to the applicable certificate of analysis that can be found at [www.atcc.org](http://www.atcc.org).

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## Growth Conditions

### Medium:

ATCC Medium 18: Trypticase Soy Agar/Broth

ATCC Medium 260: Trypticase soy agar/broth with defibrinated sheep blood

**Temperature:** 37°C

**Atmosphere:** Microaerophilic

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## Handling Procedures

1. This organism is shipped frozen in dry ice. Just prior to use, thaw vial in water at approximately 37°C. When thawed, a drop of the suspension may be used to do an immediate wet mount to observe the unique morphology of this organism and verify its viability by checking for motility.
2. Aseptically transfer the thawed suspension into a fresh #18 broth (3-5 mL). Mix

well. This suspension can now be used to inoculate agar slant(s), plate(s), or the preferred biphasic culture. Two #260 plates should be inoculated, one for microaerophilic growth and the second for aerobic growth. No growth should occur on the plate incubated aerobically.

3. To obtain a biphasic culture, add 0.6 mL of the suspension to a # 260 slant. The resulting pool at the bottom of the slant is where the best, most rapid growth will occur.
4. Incubate at 37°C under microaerophilic conditions using an anaerobe jar with an active catalyst and a microaerophilic gas generator pack, or other acceptable method, to obtain microaerophilic conditions. An oxygen concentration of 6% is ideal. Incubate tubes with cap loose.
5. Within 3 days, good growth should be obtained in the broth pool at the bottom of the slant. Additional incubation may be required for colonies to appear on agar plate. Further subcultures can be made using the broth pool as the inoculum source. Subcultures to biphasic cultures will require only 24 to 48 hours of incubation for good growth.

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## Notes

This is a slow growing organism that requires moist conditions for best growth. Growth at the broth/agar interface of the biphasic slant should occur within three days, but little turbidity will be seen. To observe growth, examine a wet mount of the broth under phase microscopy. The organism is a medium size, regular to slightly curved, motile bacillus. Motility is usually observed only in young cultures. The presence of spheroid cells indicates that viability is being lost either due to age or too much exposure to oxygen.

Once good growth is present, these organisms tend to lose viability, especially if exposed to air for lengthy periods. Viability also decreases with repeated subculturing. The cells do not Gram stain well using traditional procedures. To obtain the best results, use a basic fuchsin counterstain in place of the safranin.

Once good growth is obtained, transfer or freeze the culture. Adding an equal amount of 20% sterile glycerol to pooled broth from several biphasic slants, followed by freezing in liquid nitrogen or "ultra-low temperature" freezer is recommended.

Additional information on this culture is available on the ATCC® web site at [www.atcc.org](http://www.atcc.org).

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## Material Citation

If use of this material results in a scientific publication, please cite the material in the following manner: *Helicobacter pylori* (Marshall et al.) Goodwin et al. (ATCC BAA-945)

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## References

References and other information relating to this material are available at [www.atcc.org](http://www.atcc.org).

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## Revision

# ***Helicobacter pylori* (Marshall et al.) Goodwin et al.**

**BAA-945**

Product Sheet

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