STC-1

CRL-3254™

Description

STC-1 is an intestinal neuroendocrine cell line that was isolated in 1990 from the intestine of a 10-to-13-week-old mouse with carcinoma. The STC-1 cell line was derived from the intestinal tumors of double transgenic mice. Transgenic mice harboring a hybrid gene linking the rat insulin promoter (RIP) to polyoma small T (PyST) antigen were mated with transgenic mice harboring rat insulin promoter (RIP) linked to SV40 early region (Tag) creating off-spring harboring both transgenes (double transgenics). These mice were found to have frequent intestinal tumors in addition to pancreatic Beta-cell tumors. Gene expression studies suggested that the intestinal and pancreatic tumors arose as separate entities. The STC-1 cell line produces the hormone secretin. This cell line may be a useful model for human endocrine neoplasms of the gut. This cell line was deposited by Douglas Hanahan, PhD Cold Spring Harbor Laboratory.

Organism: *Mus musculus*, mouse
Cell Type: intestinal neuroendocrine cell
Tissue: Intestine
Age: 10 to 13 weeks
Morphology: epithelial-like
Growth properties: Adherent
Disease: Carcinoma; Invasive Small Intestinal Neuroendocrine
Cells per vial: 1.0 x 10^6
Volume: 1.0 mL

Storage Conditions

Product format: Frozen
Storage conditions: Vapor phase of liquid nitrogen

Intended Use

This product is intended for laboratory research use only. It is not intended for any animal or human therapeutic use, any human or animal consumption, or any diagnostic use.

BSL 1

ATCC determines the biosafety level of a material based on our risk assessment as guided by the current edition of *Biosafety in Microbiological and Biomedical Laboratories (BMBL)*, U.S. Department of Health and Human Services. It is your responsibility to understand the hazards associated with the material per your organization's policies and procedures as well as any other applicable regulations as enforced by your local or national agencies.

ATCC highly recommends that appropriate personal protective equipment is always used when handling vials. For cultures that require storage in liquid nitrogen, it is important to note that some vials may leak when submersed in liquid nitrogen and will slowly fill with liquid nitrogen. Upon thawing, the conversion of the liquid nitrogen back to its gas phase may result in the vial exploding or blowing off its cap with dangerous force creating flying debris. Unless necessary, ATCC recommends that these cultures be stored in the vapor phase of liquid nitrogen rather than submersed in liquid nitrogen.

Certificate of Analysis

For batch-specific test results, refer to the applicable certificate of analysis that can be found at www.atcc.org.

Growth Conditions

Temperature: 37°C
Atmosphere: 95% Air, 5% CO_2

Handling Procedures

Unpacking and storage instructions:

1. Check all containers for leakage or breakage.
2. Remove the frozen cells from the dry ice packaging and immediately place the cells at a temperature below -130°C, preferably in liquid nitrogen vapor, until ready for use.

Complete medium: The base medium for this cell line is ATCC-formulated Dulbecco's Modified Eagle's Medium, Catalog No. 30-2002. To make the complete growth medium, add the following components to the base medium: fetal bovine serum to a final concentration of 10%.

Handling Procedure:

To insure the highest level of viability, thaw the vial and initiate the culture as soon as possible upon receipt. If upon arrival continued storage of the frozen culture is necessary, it should be stored in liquid nitrogen vapor phase and not at -70°C. Storage at -70°C will result in loss of viability.

1. Thaw the vial by gentle agitation in a 37°C water bath. To reduce the possibility of contamination, keep
the O-ring and cap out of the water. Thawing should be rapid (approximately 2 minutes).
2. Remove the vial from the water bath as soon as the contents are thawed, and decontaminate by
dipping in or spraying with 70% ethanol. All of the operations from this point on should be carried out
under strict aseptic conditions.
3. Transfer the vial contents to a centrifuge tube containing 9.0 mL complete culture medium and spin at
approximately 125 x g for 5 to 7 minutes.
4. Re-suspend cell pellet with the recommended complete medium (see the specific batch information for
the culture recommended dilution ratio) and dispense into a 25 cm^2 or a 75 cm^2 culture flask. It is

important to avoid excessive alkalinity of the medium during recovery of the cells. It is suggested that,
prior to the addition of the vial contents, the culture vessel containing the complete growth medium be
placed into the incubator for at least 15 minutes to allow the medium to reach its normal pH (7.0 to 7.6).
5. Incubate the culture at 37°C in a suitable incubator. A 5% CO_2 in air atmosphere is recommended if
using the medium described on this product sheet.

This item is original depositor material that came in originally from CSHL as a safe deposit item and is repackaged, ATCC has not produced any of this material here, the material has been only QC tested. Therefore, we can only provide our general instructions as above.

Subculturing procedure: Cells must be subcultured when they reach ~70% confluence, or else they start to come off the flask into suspension.
Volumes used in this protocol are for 75 cm2 flasks; proportionally reduce or increase amount of dissociation medium for culture vessels of other sizes.

1. Remove and discard culture medium. Briefly rinse the cell layer with Ca++/Mg++ free Dulbecco's phosphate-buffered saline (D-PBS) (ATCC 30-2200) or 0.05% Trypsin – 0.02% EDTA (ATCC PCS-999-003) solution to remove all traces of serum which contains trypsin inhibitor.
2. Add 2.0 to 3.0 ml of Trypsin-EDTA solution to flask and observe cells under an inverted microscope until cell layer is dispersed (usually within 5 to 15 minutes).
   Note: To avoid clumping do not agitate the cells by hitting or shaking the flask while waiting for the cells to detach. Cells that are difficult to detach may be placed at 37°C to facilitate dispersal.
3. Add 6.0 to 8.0 ml of complete growth medium and aspirate cells by gently pipetting.
4. Transfer cell suspension to a centrifuge tube and spin at approximately 125 xg for 5 to 10 minutes.

5. Discard supernatant. Resuspend the cell pellet in fresh growth medium.
6. Add appropriate aliquots of the cell suspension to new culture vessels. Incubate cultures at 37°C.

Subcultivation Ratio: 1:3 to 1:5 is recommended.
Medium Renewal: Every 2 to 3 days
Reagents for cryopreservation: Complete growth medium supplemented with 10% (v/v) DMSO (ATCC 4-X)

Material Citation

If use of this material results in a scientific publication, please cite the material in the following manner: STC-1 (ATCC CRL-3254)

References

References and other information relating to this material are available at www.atcc.org.

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